

VALENTINA DEBOLINA MONDAL

M.Sc. Microbiology

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OBJECTIVES

To work in a professional atmosphere that enables to acquire emerging trends and technologies to widen the spectrum of my knowledge, and contribute towards organization and individual growth.

EDUCATIONAL BACKGROUND

- ↑ ↓ Completed Masters in the year **2020** in Microbiology from University of Calcutta affiliated **VIJAYGARH JYOTISH RAY COLLEGE** with **6.45 CGPA** marks.
- ↑ ↓ Completed Bachelors in the year **2018** in Microbiology from University of Calcutta affiliated **SCOTTISH CHURCH COLLEGE** with **57.88 %** marks.
- ↑ ↓ Passed Higher Secondary Examination (**WBCHSE**) from **HOLY CHILD INSTITUTE** with **71%** marks in the year **2015**.
- ↑ ↓ Passed Secondary Examination (**WBBSE**) from **HOLY CHILD INSTITUTE** with **69.72%** marks in the year **2013**.

INTERSHIPS / TRAININGS

- **CSIR-IICB** Integrated Skill Development certification course on Theory and Hands-on-Training in High end equipments for clinical applications- Optical Microscopy from 22nd August 2022 to 2nd September 2022.
- **COVID Unit, Virology Laboratory, School of Tropical Medicine, Kolkata**

Sample Collection from patients, RNA Extraction (manual and automated) from Nasopharyngeal and Oropharyngeal samples, RT- qPCR, ELISA (non-COVID) under the guidance of In-charge Virology Unit, Prof. (Dr.) Bhaswati Bandyopadhyay from 31st December 2021 to 30th March 2022.

- Participated and successfully completed the **Proteomics Advanced Winter School 2021** organized by Proteomics Lab, Indian Institute of Technology Bombay during 8th – 19th November, 2021 under the guidance of Prof. Sanjeeva Srivastava.

SUMMER PROJECT

Institute : National Institute Of Cholera and Enteric Disease (NICED)

Supervisor : Dr. Nilanjan Chakraborty

Duration- June 2019 to August 2019 (2 months)

Title : MOLECULAR DIAGNOSIS OF CONGENITAL CYTOMEGALOVIRUS INFECTION
AMONG INFANT PATIENTS.

Techniques Used : Genomic DNA Isolation, Polymerase Chain Reaction, Agarose Gel Electrophoresis.

Project Highlights :

- Worked on the urine and blood samples which were collected from the infant patients from different metropolis hospitals & genomic DNA isolation were performed with the collected samples.
- Molecular diagnosis of Human Cytomegalovirus (HCMV) infection from these samples of infant patients were done using Polymerase Chain Reaction.
- Also the most prevalent symptoms were studied by comparing the collected data from these infant patients.

EQUIPMENTS HANDLED

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| ● Autoclave, Laminar Air Flow and Hot air oven | ● ELISA reader, Applied Biosystems 2720 Thermal Cycler machine |
| ● BOD Incubator and Shaker Incubator, Colony counter | ● Thermo Scientific TM KingFisher Flex Purification System |
| ● UV-Visible Spectrophotometer and Gel docs | ● BIO-RAD CFX96 Real Time PCR Machine |
| ● Microscopes, Centrifuges and Ultracentrifuges | ● Laser Scanning Confocal Microscope |

LABORATORY TECHNICAL SKILLS

- Basics (Pipette Handling, Dilutions, Microscope Handling, Spectrophotometer Handling and OD determination, Preparation of complex media & chemically defined synthetic media, Streaking on agar plate & agar slant, Pour plate method)
- Staining (Endospore, Capsular, Flagellar, Fungal, Actinomycetes and Gram Staining)
- Serial Dilution Method for Bacterial Cultures
- Sub Culturing of Microbes
- Isolation and Culturing of microbes from Air, Water and Soil samples
- SDS PAGE and Agarose Gel Electrophoresis
- Cell culture, Cell seeding, Cell transfection, Cell fixing and Imaging by indirect or direct immunofluorescence imaging using Laser Scanning Confocal Microscope followed by the image analysis using ZEN imaging software and Bitplane Imaris software
- Basic Chromatographic techniques (Thin Layer Chromatography and Paper Chromatography)
- Cell Culture and maintenance
- Competent cell preparation
- Plasmid isolation and Genomic DNA isolation, RNA Extraction
- ELISA and Immunodiffusion
- Antibiotic assay techniques and Drug Assay
- Western Blot technique
- Endpoint Polymerase Chain Reaction, Real Time RT-PCR
- Sequence Alignment (NCBI-BLAST) and Phylogenetic Tree Construction (Clustal W2)
- Primer Designing for PCR using Gene Runner
- Biochemical tests for the Screening of Microbes etc.

PARTICIPATIONS

- ❖ Poster Presentation delivered on “Microbes in our Life” organized by Department of Microbiology in association with Research and Guidance Cell of Vijaygarh Jyotish Ray College on 7th May 2019.

Topic: ‘Community Genomic Analysis Dissimilatory Arsenate Reducing Bacteria in Aquifer Sediments of Bengal Delta Plain’ organised by Vijaygarh Jyotish Ray College, Kolkata.

PERSONAL DETAILS

D.O.B.- 31st August, 1996

Father's Name- David Sanjoy Mondal

Nationality- Indian

REFERENCES

- Prof. (Dr.) Bhaswati Bandyopadhyay, In-charge Virology Unit,
Department of Microbiology, Calcutta School Of Tropical Medicine.
Kolkata, INDIA 700073
Email: drbhaswati@yahoo.com
- Dr. Nilanjan Chakraborty, Scientist F Division,
Virus Research Laboratory, ICMR-NICED.
Kolkata, INDIA 700010
Email: nilanjan_19@yahoo.com
- Dr. Prasenjit Das, Professor Head,
Microbiology Department, Vijaygarh Jyotish Ray College.
Kolkata, INDIA 700032
Email: pdmicrobio@gmail.com

LANGUAGE PROFICIENCY

Native or bilingual proficiency in Bengali, full professional proficiency in Hindi and English.

COMPUTER PROFICIENCY

- Microsoft Office (Word, Powerpoint, Excel) and basics in Computer Application
- Adobe Illustrator

EXTRA CURRICULARS Drawing and Photography.

DECLARATION: I do hereby declare that the above information is true to my belief and knowledge.

PLACE : KOLKATA, INDIA.

Valentina Debolina Mondal

SIGNATURE

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